

Lecture #9

Fundamentals of microfluidics and microfluidic chips

Aims:

- Understand the behavior of liquids at the microscale
- Learn the basics about continuous flow and droplet microfluidics
- Get an overview on the basic chip manufacturing principles
- Get an overview on the different droplet modules and functionalities
- Prominent applications

Lectures (CO 121)	Date & Topic	Details	Practical (location as color coded on next slide)
1	13.09 General Intro	Get to know teachers, TAs, students and aims of the course	17.09 Measure temperature using thermistor (using M&A explorer) TL
2	20.09 Lecture LabVIEW TL Group formation (A-F, 3 students, each)	Some first basic steps in LabVIEW programming	24.09 Brief intro into LabVIEW thermistor program (input and output) TL
3	27.09 Case study FACS, similarities and differences to droplet microfluidics Selection of case study topics	1.) Property to measure? 2.) Device? 3.) Working principle? 4.) Alternatives?	01.10 Preparation of bioinstrument case study
4	04.10 No course, preparation for case study		08.10 No course
5	11.10 Groups A-B presenting case study		15.10 LBMM workstation labs, intro into Nature Protocols
6	18.10 Lecture optics Homework: Students to prepare one laser/PMT blueprint FP	Mirrors, filters, microscopes, lenses, etc.	22.10 Holidays
	25.10 Holidays, submit your blueprint by email		29.10 .10 Build workstation optics 1
7	01.11 Lecture electronics	FPGA, PMTs, amplifier, function generator	05.11 Build workstation 1 optics 2, laser alignment; build workstation electronics
8	08.11 Intro into enzyme kinetics and measurement experiment (kinetics, measurement)	Enzymes, kinetics, practical task	12.11 -
9	15.11 -	Software similar to Thermistor program, pdf on installation	19.11 Intro to droplet analysis software (LabVIEW) TL Build workstation software: Add output LED (mimicking sorting trigger) into analysis software
10	22.11 Fundamentals of microfluidics and microfluidic chips	Flow at the microscale, microfluidic chips (manufacturing), droplet microfluidic modules	26.11 Run microfluidic experiments, e.g. determine concentration of MMP in droplets
11	29.11 Prepare presentation		3.12 Sorting Demo on LBMM workstation1 (Groups A-B)
12	06.12 Prepare presentation		10.12
13	13.12 Groups B-A presenting results 13.12 Submit report (all!)		17.12 – TUESDAY! - Individual Q & A sessions (10min, Groups A-B)

Last lecture before YOUR microfluidics experiment!

Green shading: Single seminar/practical with all 18 students
Red shading: Individual seminar/practical with only 6 students required (= 3 sequential 90min slots, 4.5h in total)

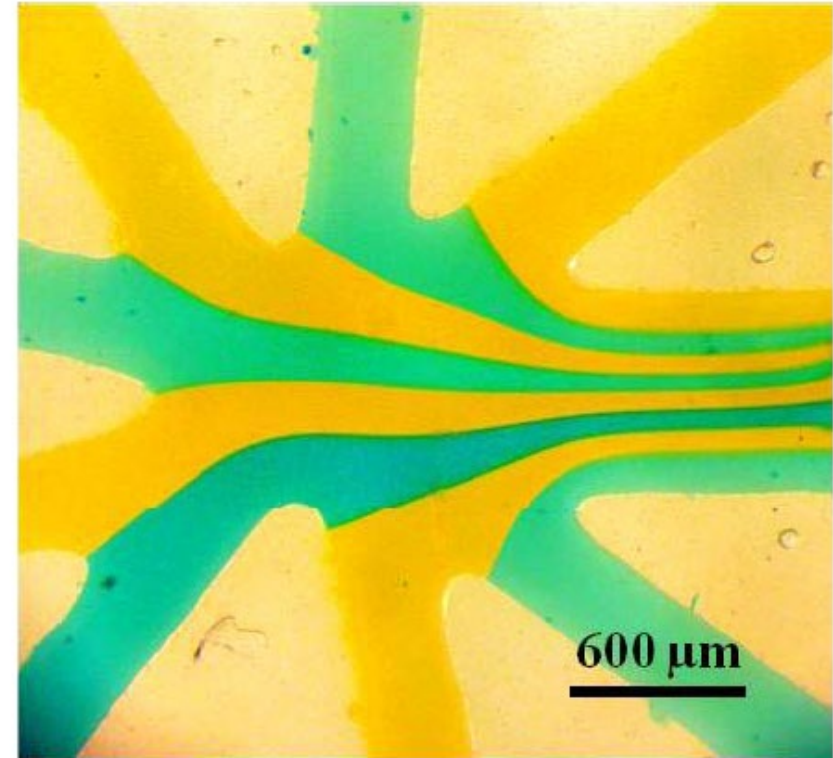
A general difference between macroscopic and microscopic flow

What is happening here?



<https://www.azom.com/article.aspx?ArticleID=14131>

Macroscopic flow is turbulent



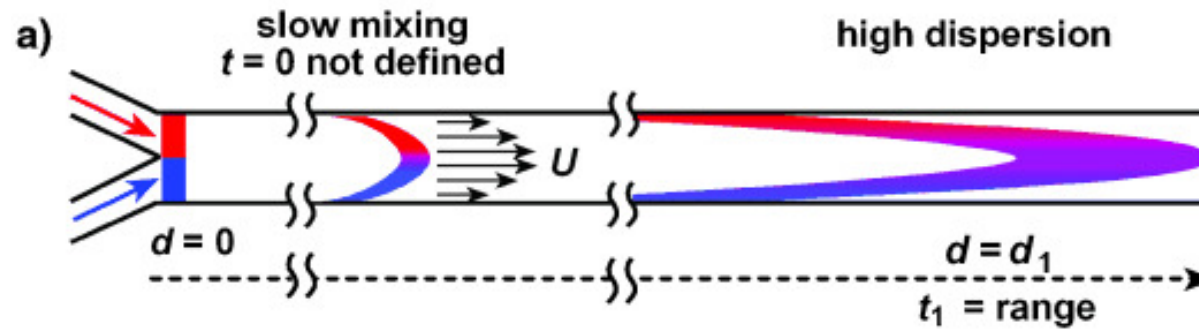
http://alcheme.tamu.edu/?page_id=6720

Microscopic flow is laminar

A general difference between macroscopic and microscopic flow

*“Consider the act of mixing sugar cubes in tea: waiting for the sugar to diffuse and mix is too slow, and rapid mixing is achieved by stirring which induces turbulent flow (convectioanal mixing). If the tea is replaced by a viscous liquid, stirring becomes harder, because viscosity damps any motion reducing turbulence and diffusive mixing dominates. **In microfluidic devices, viscous forces dominate.** But instead of changing the liquid, the same effect is obtained by reducing size. Scaling a tea cup to less than a millimeter gives an approximation of the fluid physics in a microfluidic system. **This fluid flow regimen in microfluidics is laminar and not turbulent.**”*

A general difference between macroscopic and microscopic flow



Small channel diameters imply that fluid moves very close to the static walls
=> friction and viscosity become more relevant

Reynolds number – is the flow laminar or turbulent?

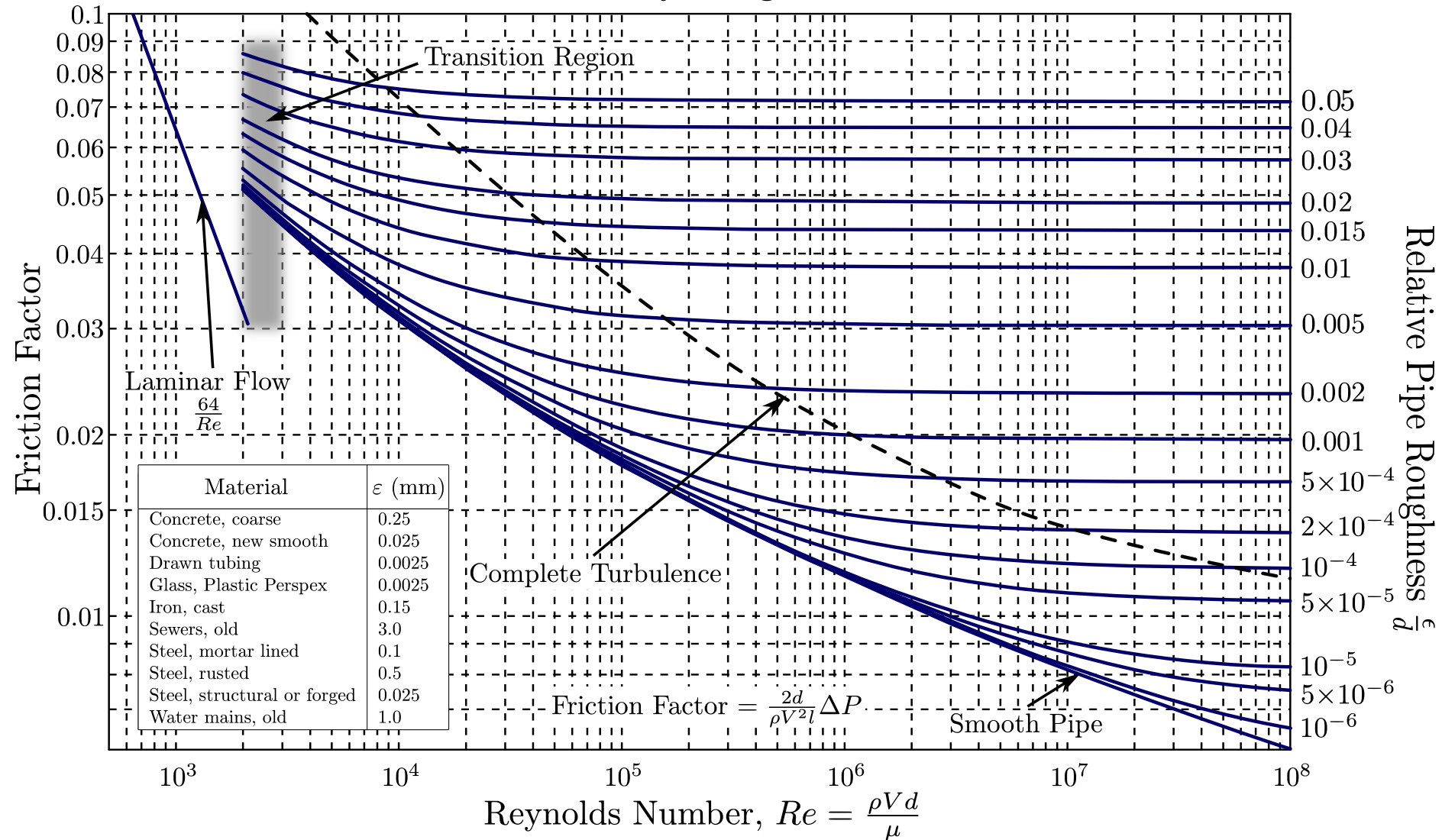
$$Re = \frac{\rho V L}{\mu}$$

With ρ = density of the fluid, V = velocity of the fluid, L = length of the channel and μ = viscosity of the liquid.

For $Re < 2300$, flow is laminar, above 4000 it becomes entirely turbulent. Exact numbers depending on roughness of channel wall (see Moody diagram)

Reynolds number – is the flow laminar or turbulent?

Moody Diagram



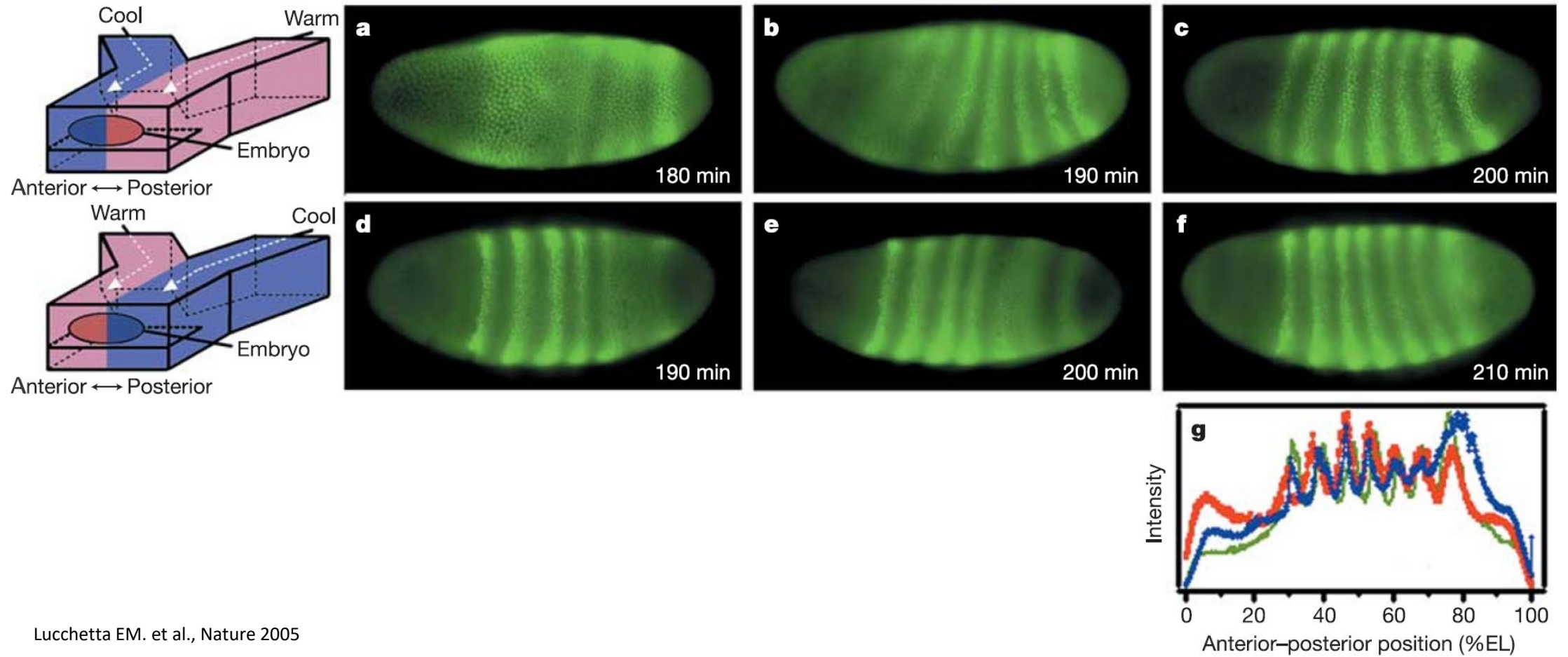
Peclet number – what drives mixing?

$$Pe = \frac{VL}{D}$$

where D is the diffusion constant, V the velocity of the fluid and L the length of the channel.

For low Pe (<1), mixing is only determined by diffusion, so very slow. This enables very special experimental setups:

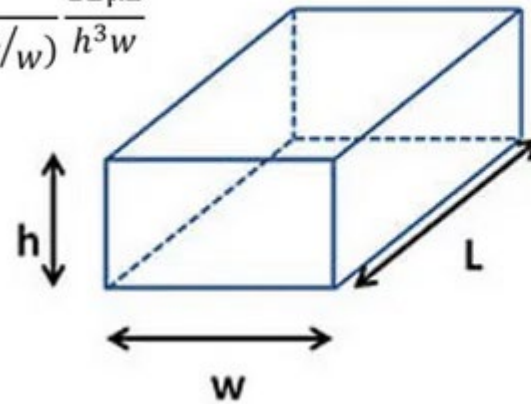
Application: Measuring the effect of temperature on embryonic development, discovery of compensation mechanisms



Other relevant parameters in continuous flow at the microscale - resistance

Rectangular cross section

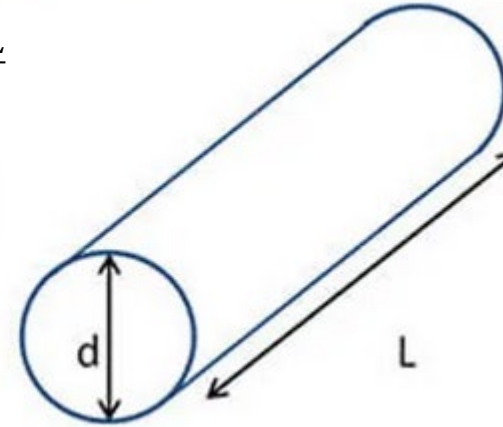
$$R = \frac{1}{1 - 0,63(h/w)} \frac{12\mu L}{h^3 w}$$



L = length of the channel ($L \gg w$)
h = height of the channel
w = width of the channel ($w \gg h$)
 μ = dynamic viscosity

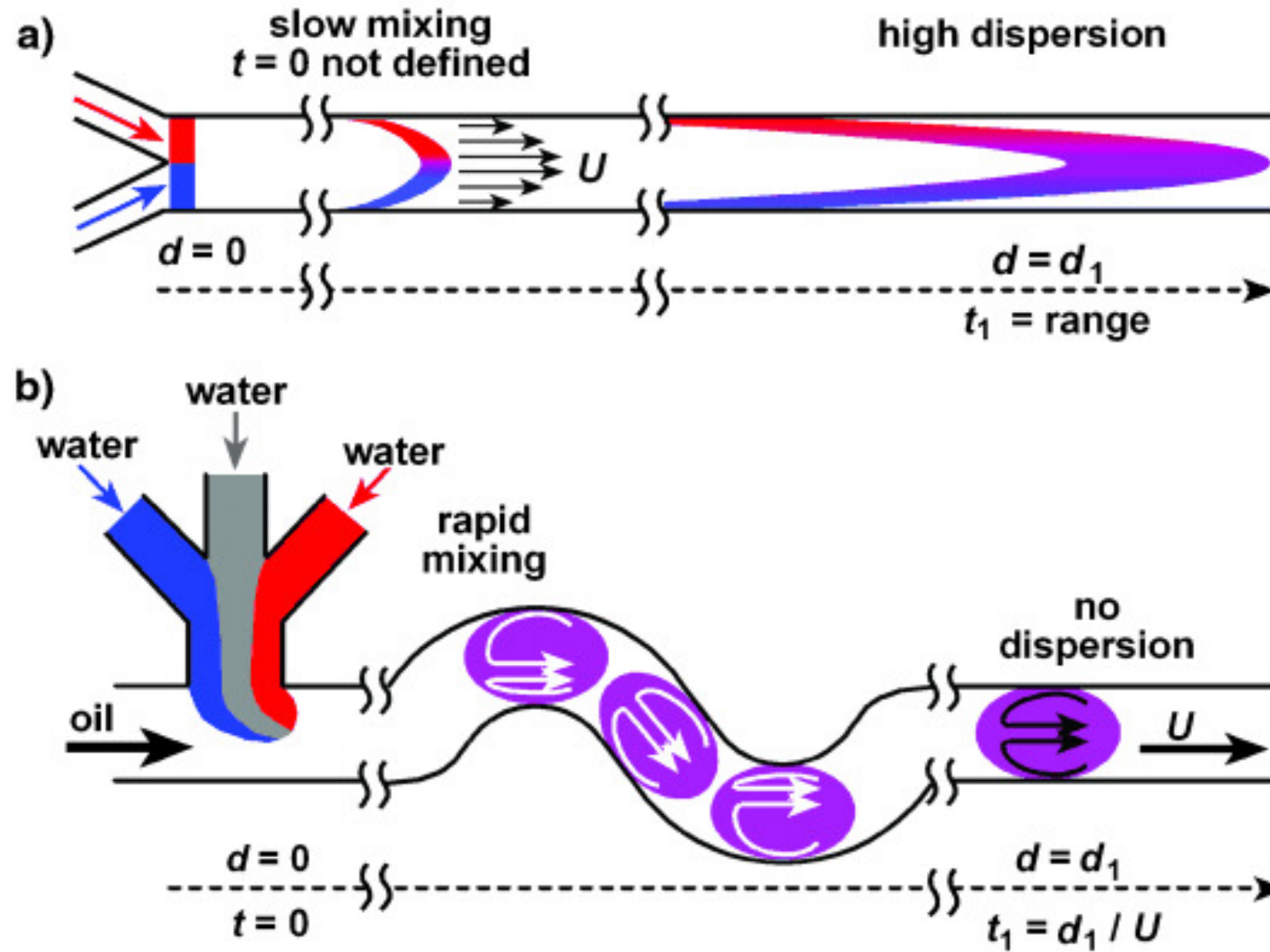
Circular cross section

$$R = \frac{128\mu L}{\pi d^4}$$



L = length of the channel ($L \gg d$)
d = diameter of the channel
 μ = dynamic viscosity

How do things change in multiphase systems?



Capillary number – stable or unstable droplets?

The capillary number Ca describes the ratio between viscous forces and surface tension between two immiscible liquids (e.g. water and oil):

$$Ca = \frac{\mu V}{\gamma}$$

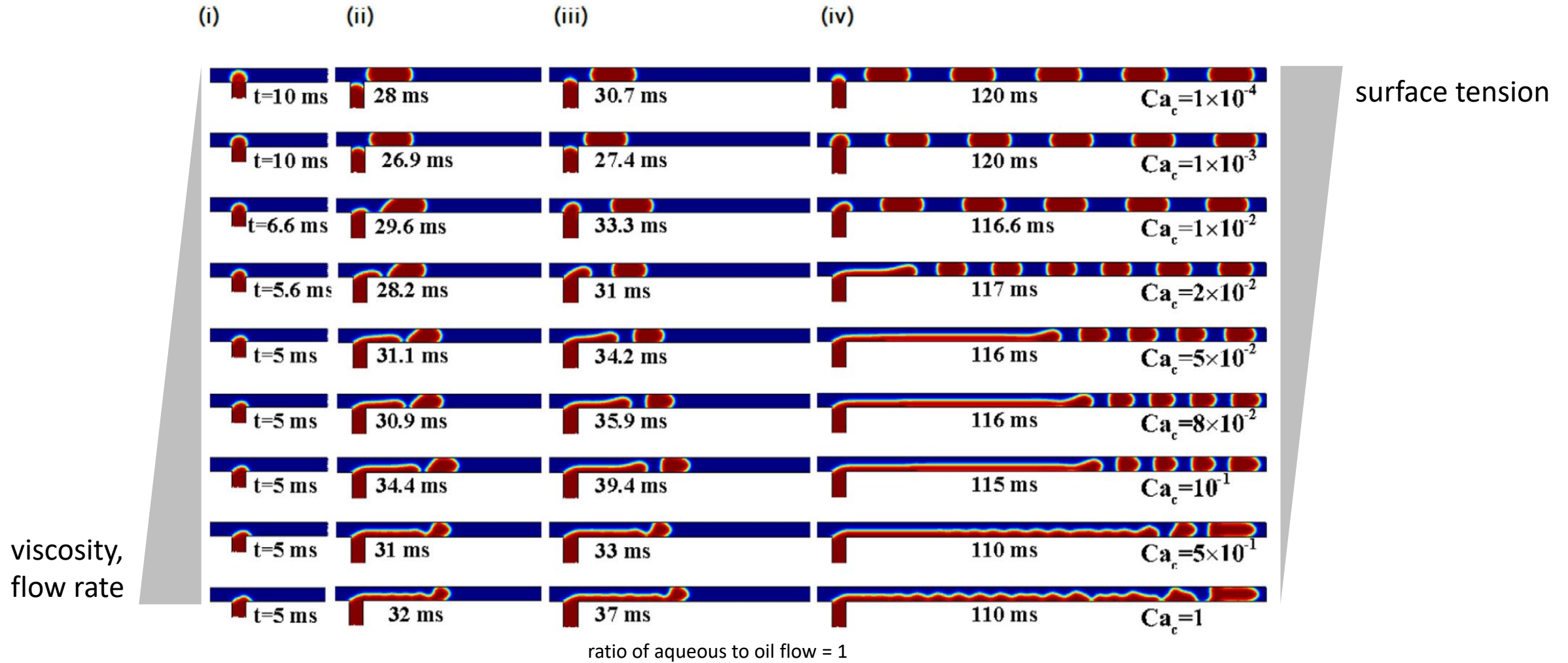
μ = viscosity of the dispersed liquid

V = velocity

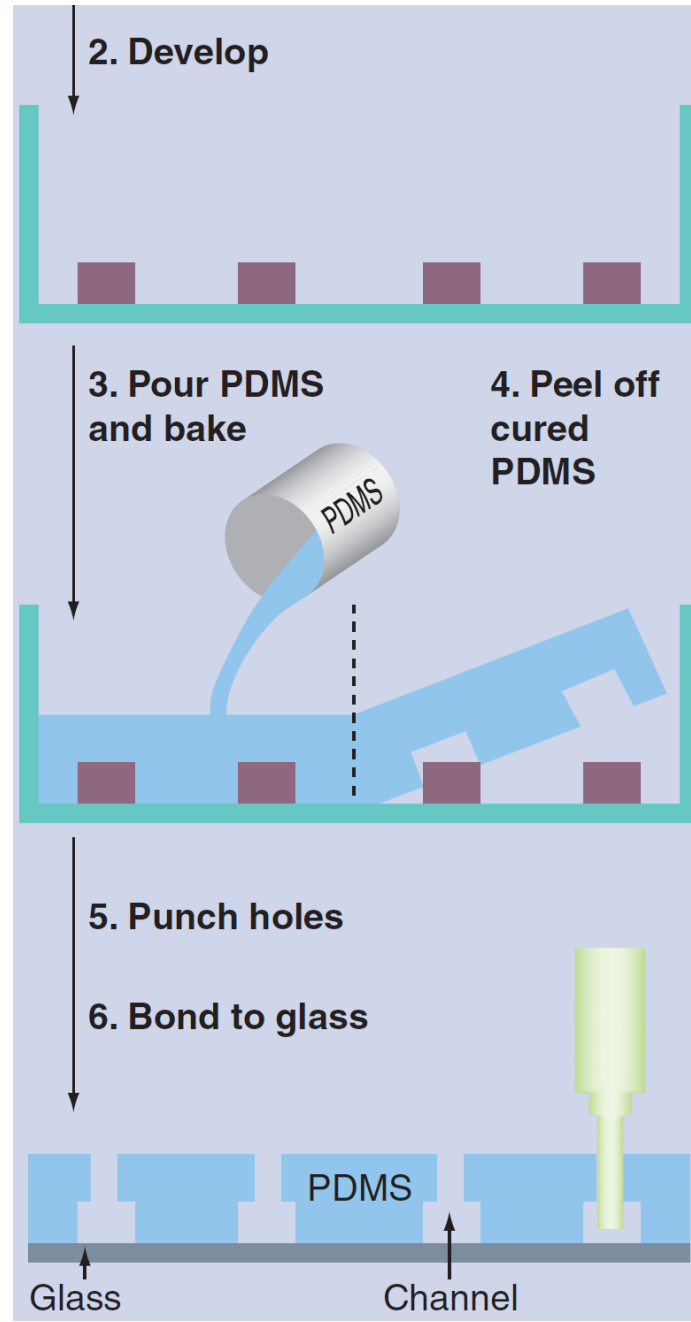
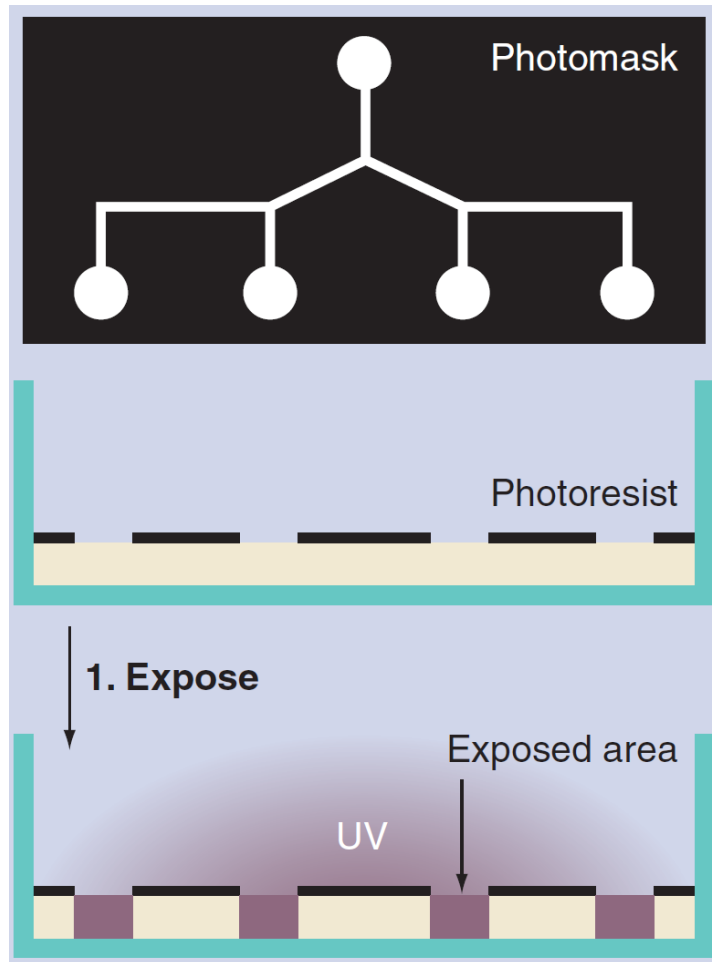
γ = surface tension between two fluid phases

In microfluidics the capillary number is usually between $10^{-3} - 10^{-1}$

Capillary number – stable or unstable droplets?

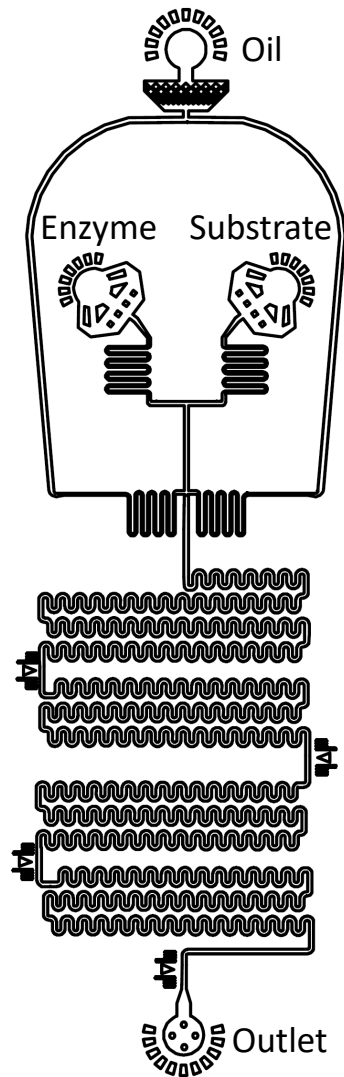


How to manufacture microfluidic chips?

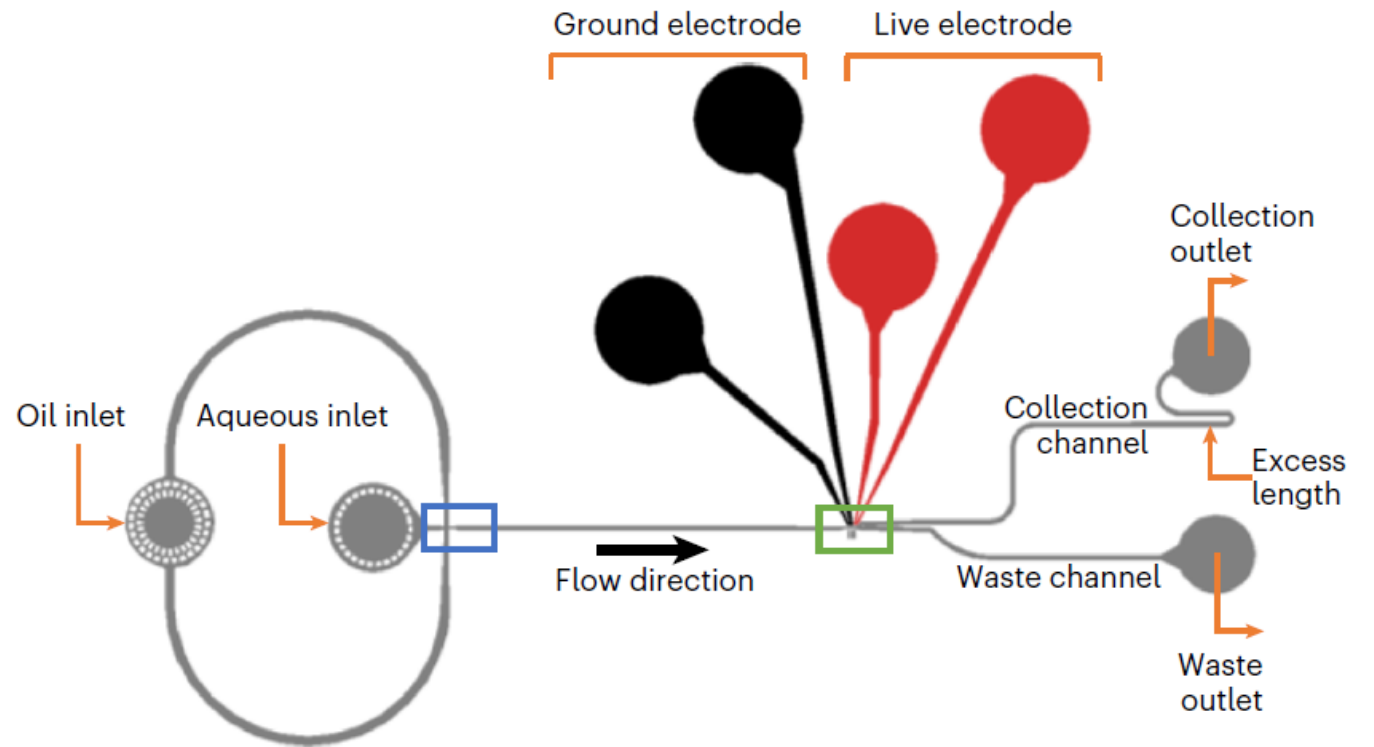


BIOENG-421 modules

Determination of enzyme concentrations (26.11.2024)



LBMM sorting demo (3.12.2024)



Some practical benefits of microfluidic systems

- Large surface to volume ratio = **very rapid heat exchange** (on-chip PCR, etc.)
- Very small volumes enabling to obtain **detectable concentrations of analytes from single cells** (single cell sequencing, phenotypic antibody screens, directed evolution, etc.)
- Low volumes reduce reagent cost and facilitate the **use of very limited material such as patient cells**
- Droplets as assay compartments (“miniaturized test tubes”) can be generated at kHz frequencies, **enabling uHTS**

Single cell RNAseq

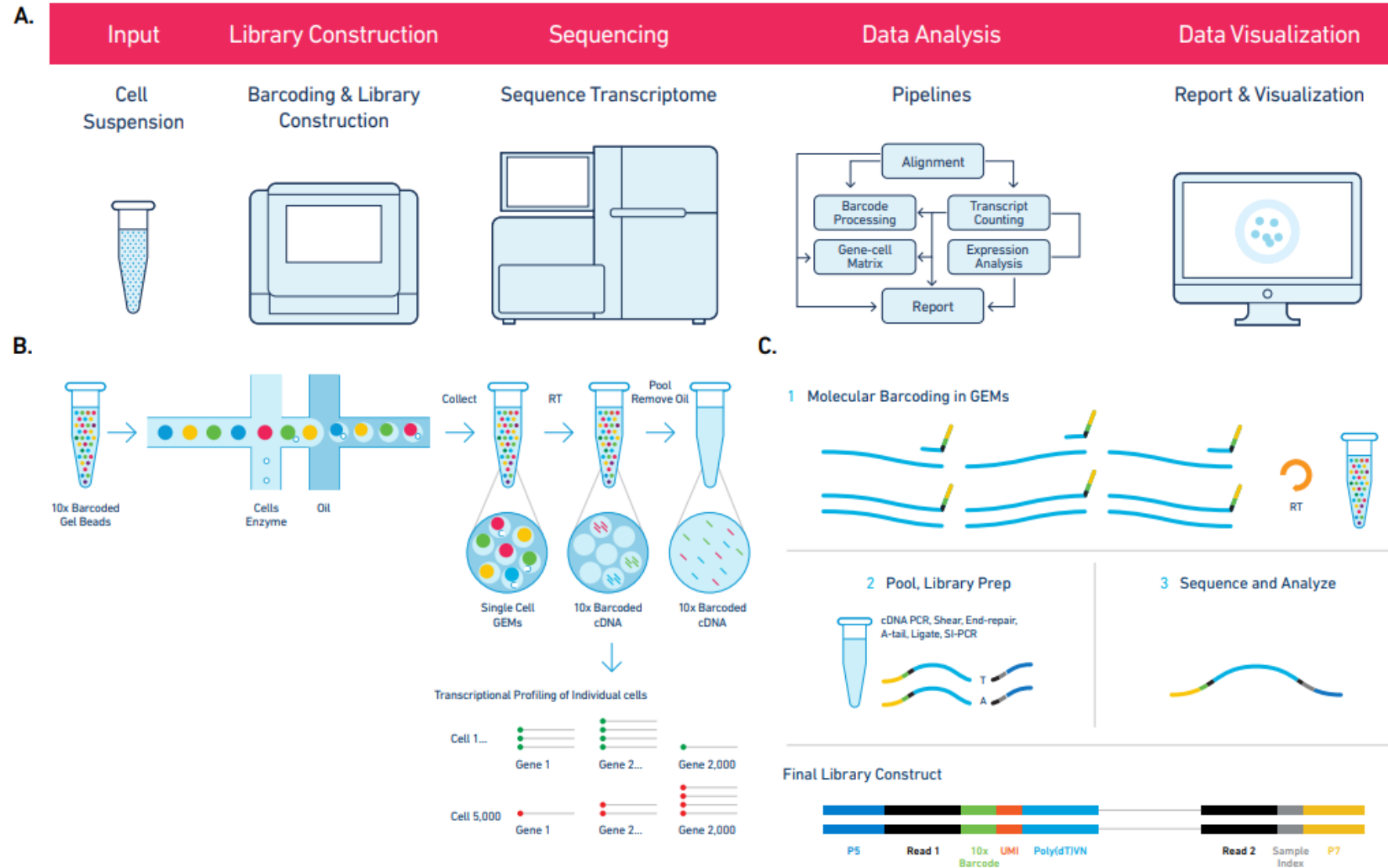
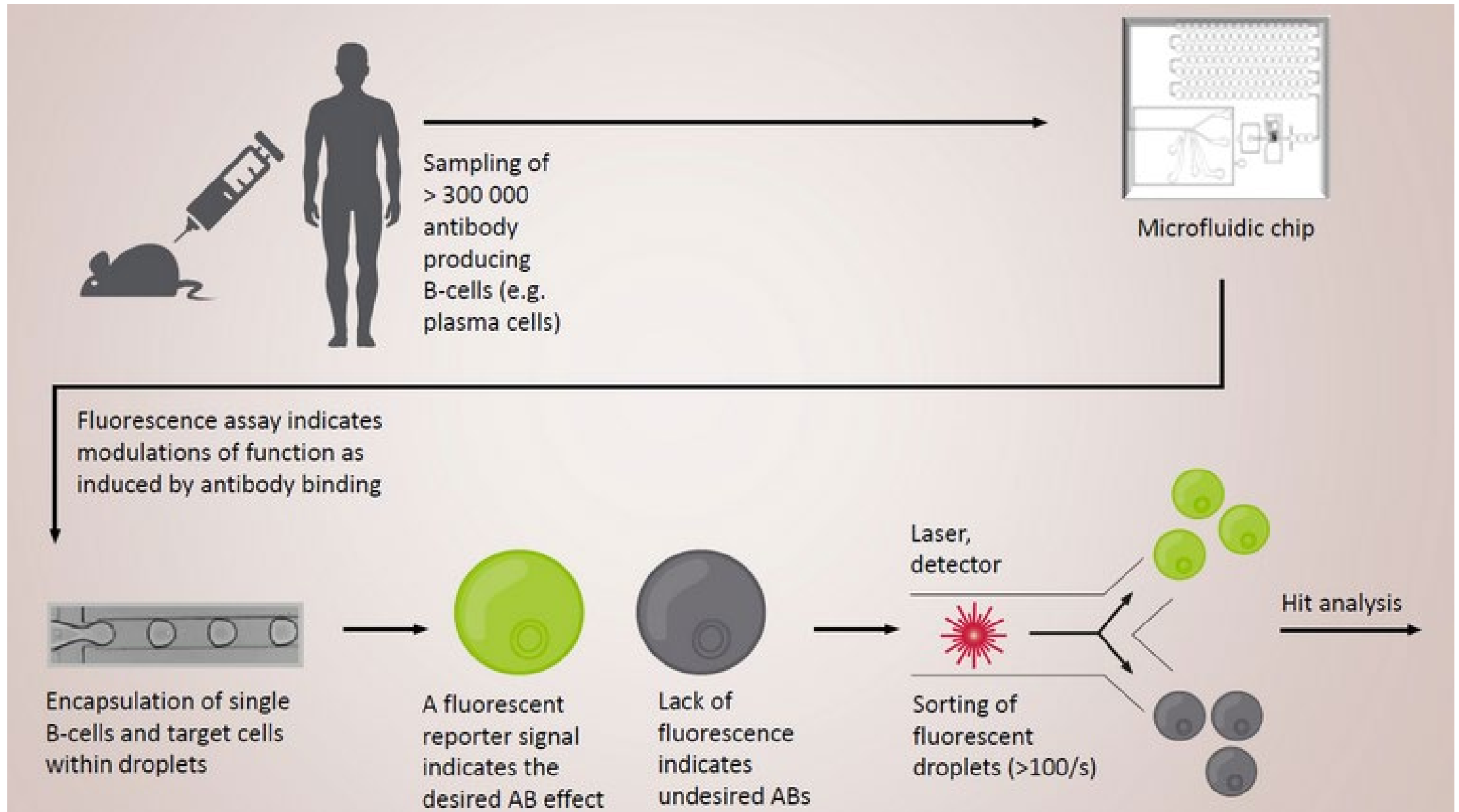
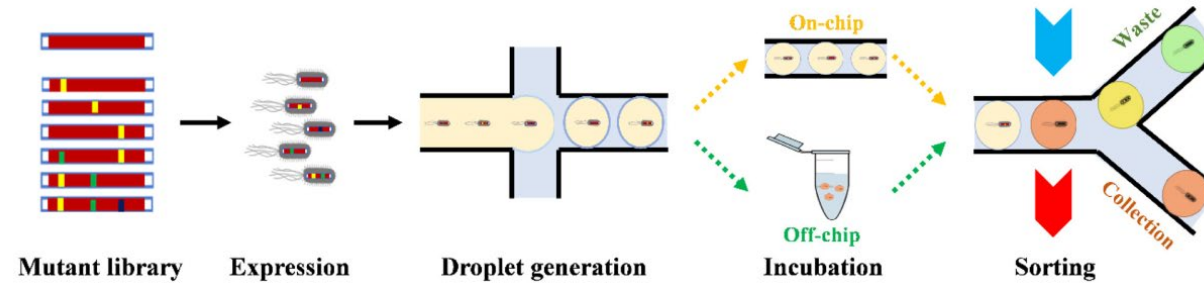


Figure 1. Chromium™ Single Cell 3' Solution. (a) Workflow schematic overview. (b) Formation of GEMs, RT takes place inside each GEM, which is then pooled for cDNA amplification and library construction in bulk. (c) v2 Single Cell Assay schematic overview.

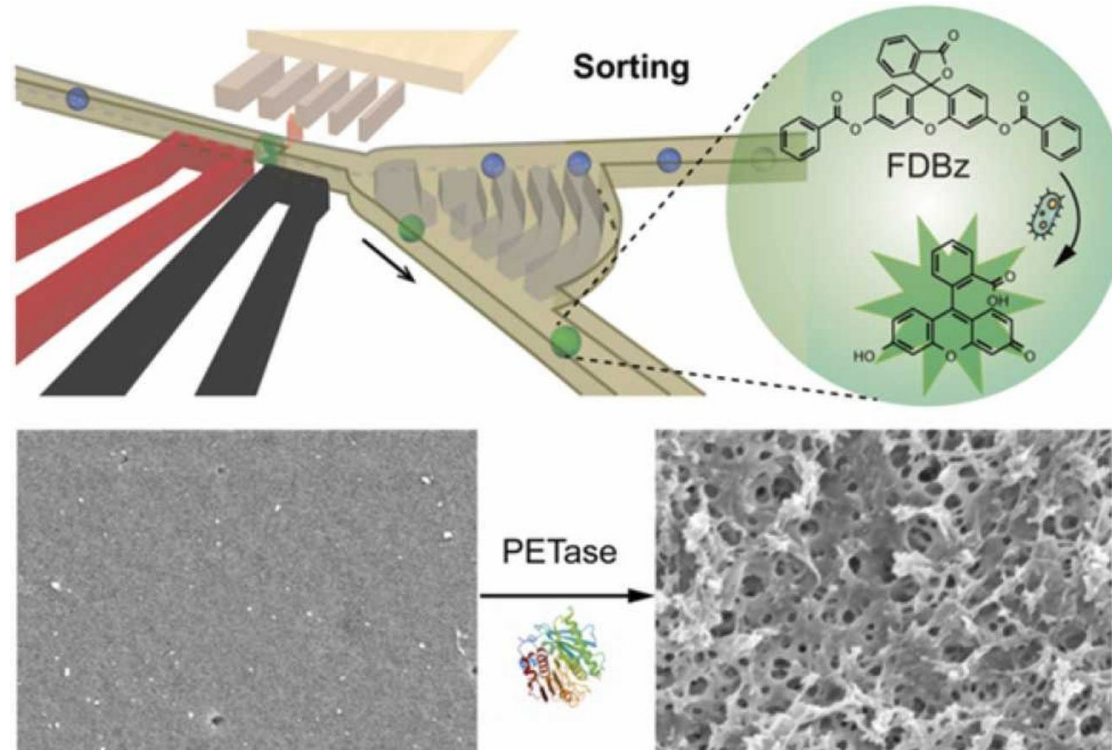
Single cell antibody screening



Directed evolution – new or improved enzyme activities

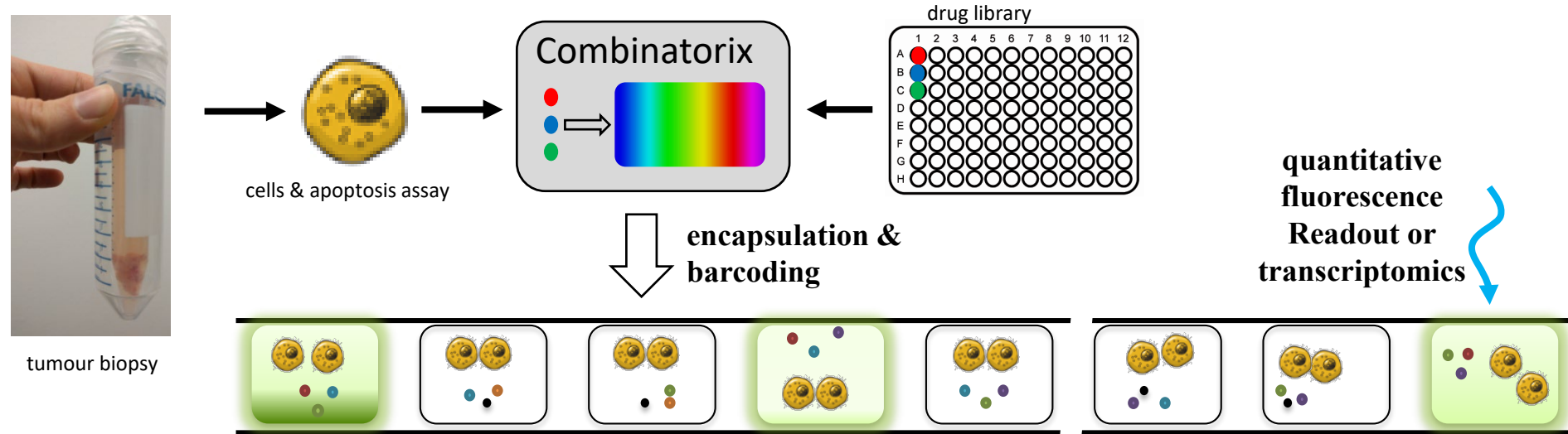


Fu et al., Front. Chem. 9:666867. doi: 10.3389/fchem.2021.666867



Qiao et al., 2022, <https://doi.org/10.1016/j.jhazmat.2021.127417>

Screening patient samples

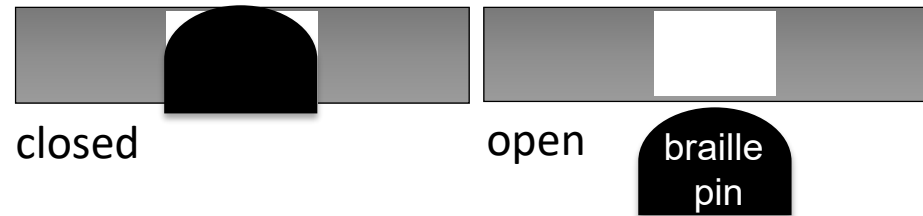


Based on the miniaturized assay volumes, one can screen about 100 times more treatment conditions on very limited patient material (e.g. cells from a biopsy) as compared to conventional formats

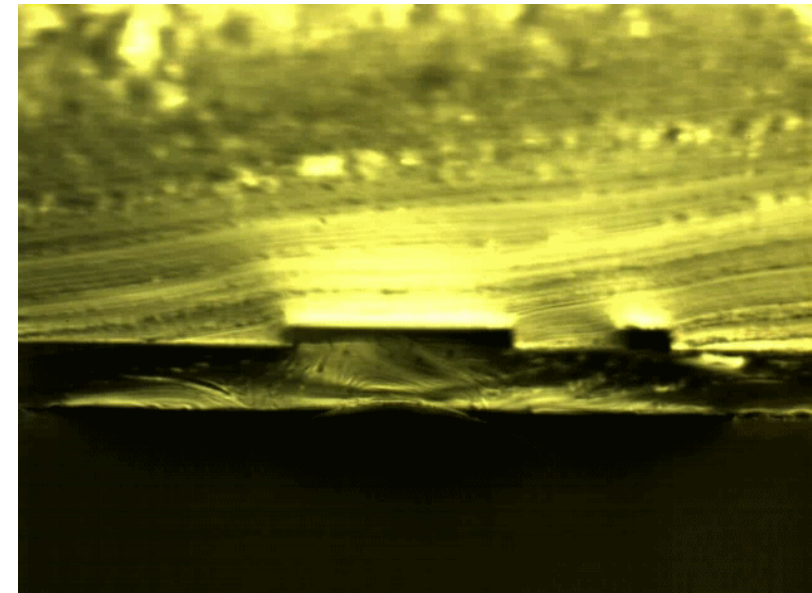
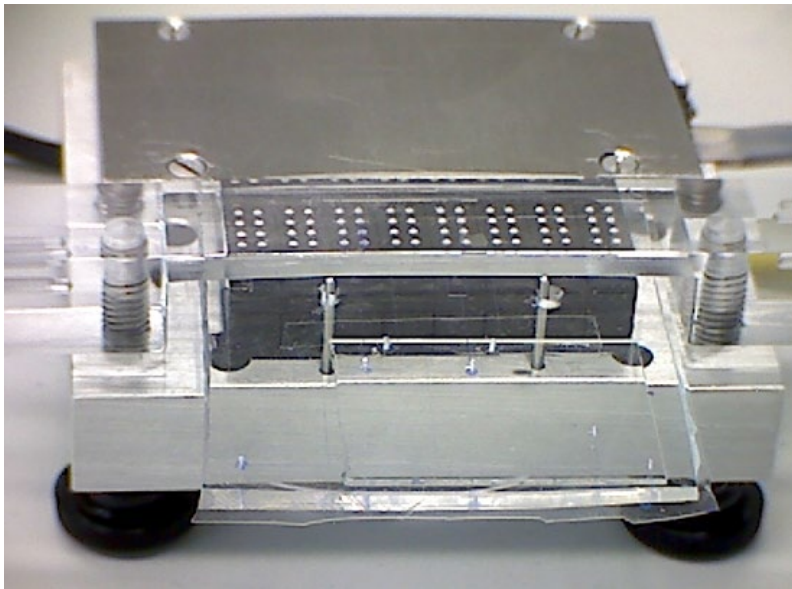
Underlying technology



Underlying technology



Ramesh
Utharala

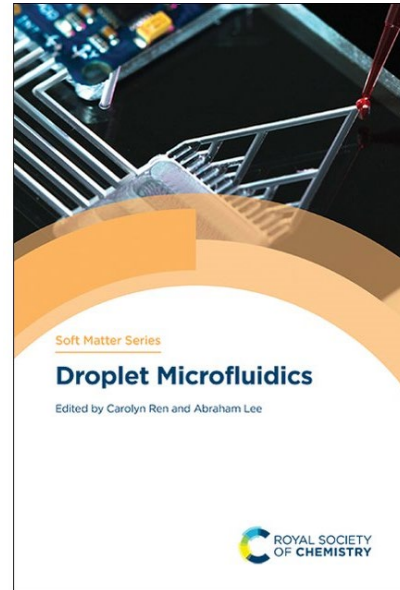
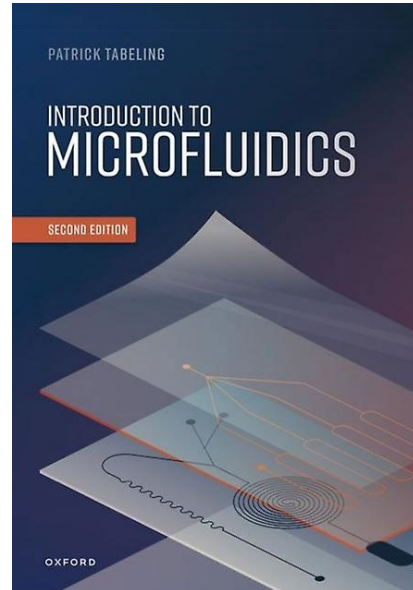


Initially conceived by Gu et al., <https://www.pnas.org/doi/10.1073/pnas.0404353101>

Microfluidics is a big playground at the interface of engineering and biology!



Further reading



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Droplet-based microfluidics

[Thomas Moragues](#), [Diana Arguijo](#), [Thomas Beneyton](#), [Cyrus Modavi](#), [Karolis Simutis](#), [Adam R. Abate](#), [Jean-Christophe Baret](#), [Andrew J. deMello](#) , [Douglas Densmore](#) & [Andrew D. Griffiths](#)

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Questions?

